SALUS PRO PATRIA

Nutritional composition of *Tenebrio molitor* larvae reared on substrates derived from by-products

Introduction

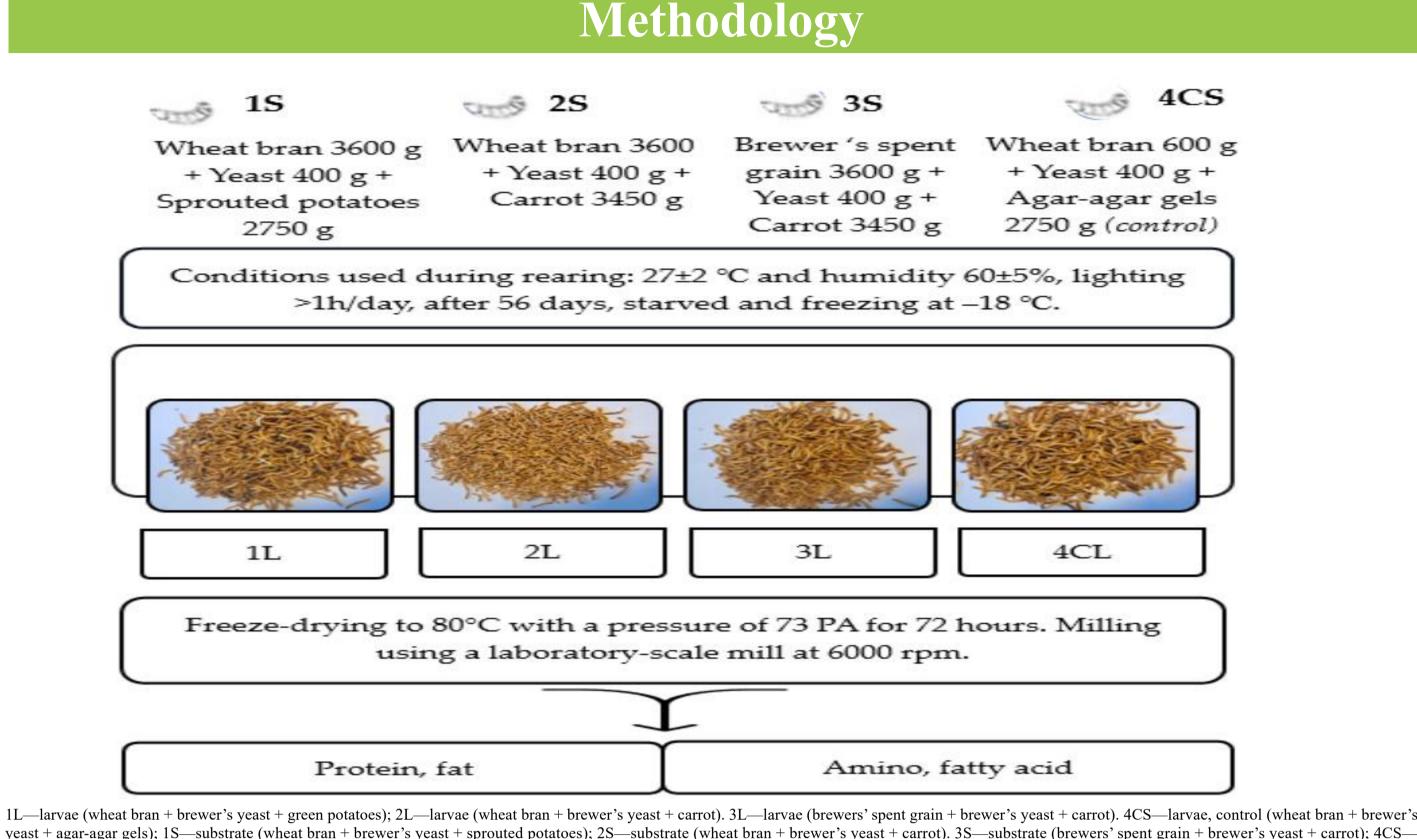
- It is projected that the world's population will reach nearly 9.8 billion people By 2050.
- Consequently, environmental sustainability concerns underscore the importance of developing alternative protein strategies, including new sources of high-quality proteins and fats. When rearing mealworms, it should be taken into account that they are grown quickly with the lowest costs and the resulting biomass is of high quality.
- The rearing of *Tenebrio molitor* larvae (Linnaeus, 1758), a member of the darkling beetle family (*Tenebrionidae*), using plant-based byproducts offers multiple benefits.
- Together with a shift towards an economical, safe, and sustainable outcome, there is an increasing trend of attempting to rear mealworms using by-products from production and agriculture.
- The impact of these by-products on the mealworms themselves can vary, depending on the chosen raw materials and their proportions.
- Analyzing the nutritional value of mealworms, besides the quantity of proteins and fats, it's crucial to consider the specific AAs and FAs present.

This study **aims** to explore the potential of using local by-products or food residues as substrates in the mass production of *Tenebrio molitor* larvae (mealworms), focusing on achieving an optimal amino and fatty acid profile.

The **object** of this study encompasses the use of various plant-based by-products such as brewers' spent grain, wheat bran, and notably, sprouted potatoes, which are considered waste from farms and are no longer suitable for human consumption.



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substrate, control (wheat bran + brewer's yeast + agar-agar gels);

Figure 1. Chart illustrating the complete research procedure.

Results

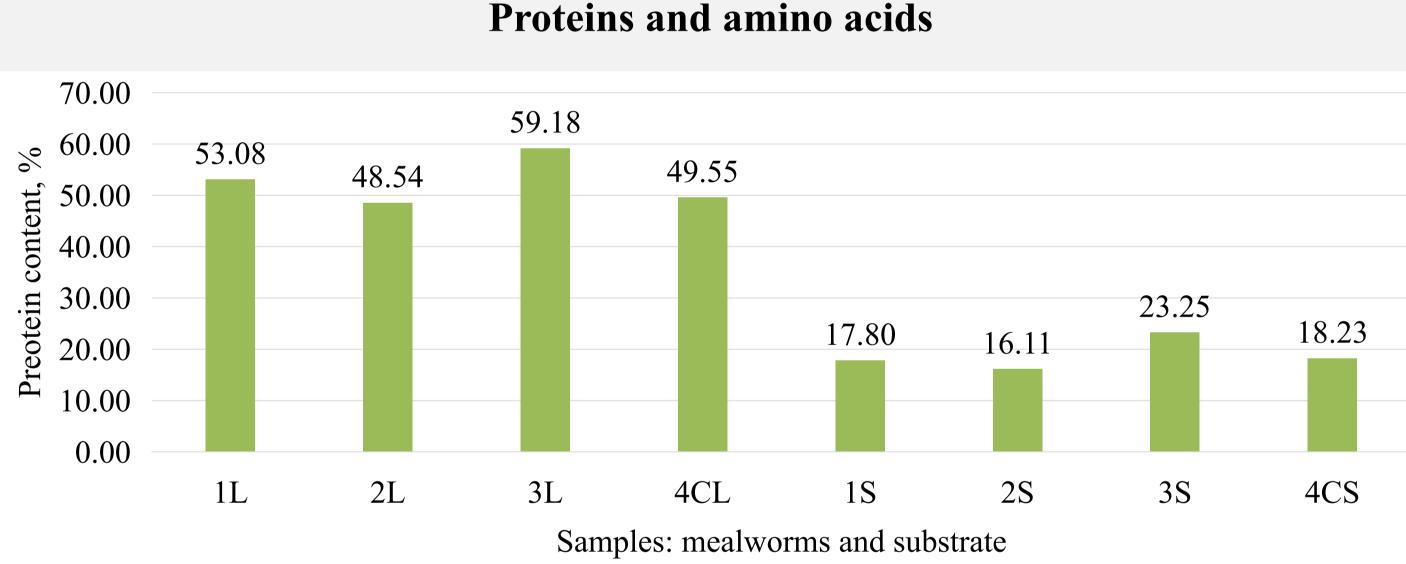


Figure 2. Protein content in substrate and mealworms

Table 1. Amino acid composition in lyophilized larvae and substrate, g/100g of dry matter, average \pm standard error, n=3

		Lar	vae		Substrate					
Samples	1L	2L	3L	4CL (control)	1S	2S	38	4CS (control)		
Valine	2.88±0.134 a	2.25±0.013 b	3.14±0.130 a	2.93±0.174 a	0.84±0.089 a	0.74±0.061 a	1.19±0.025 b	0.83±0.038 a		
Leucine	3.54±0.130 a	3.02±0.002 b	3.79±0.183 a	3.45±0.262 ab	1.09±0.060 a	1.00±0.071 a	1.53±0.009 b	1.10±0.041 a		
Isoleucine	2.01±0.104 a	1.61±0.011 b	2.14±0.096 a	2.02±0.111 a	0.63±0.085 ac	0.47±0.059 b	0.79±0.007 c	0.52±0.042 ab		
Threonine	1.39±0.082	1.27±0.045	1.45±0.002	1.34±0.111	0.45±0.004 a	0.39±0.019 b	0.52±0.013 c	0.41±0.017 b		
Methionine	0.60±0.027 a	0.46±0.044 b	0.62±0.023 a	0.57±0.026 ab	0.18±0.011 a	0.24±0.001 b	0.30±0.017 c	0.26±0.012 b		
Phenylalanine	1.63±0.065 ab	1.63±0.039 ab	1.83±0.097 a	1.61±0.089 b	0.72±0.034 a	0.66±0.045 a	1.11±0.010 b	0.69±0.034 a		
Lysine	2.73±0.060 a	2.46±0.016 b	2.92±0.152 a	2.80±0.075 a	0.80±0.048 a	0.87±0.044 ab	1.07±0.001 c	0.95±0.042 b		
Histidine	1.60±0.116 abc	1.49±0.032 b	1.81±0.116 cd	1.89±0.009 d	0.83±0.005 a	0.61±0.021 b	0.71±0.008 c	0.67±0.002 d		
Aspartic acid	3.92±0.132 a	3.21±0.035 b	4.30±0.257 a	3.78±0.306 ab	1.22± 0.056 a	1.10±0.051 a	1.47±0.048 b	1.19±0.060 a		
Glutamic acid	5.88±0.238 a	3.73±0.121 b	6.49±0.903 a	6.17±0.046 a	3.86±0.254 a	2.80±0.199 b	3.67±0.171 a	3.00±0.081 b		
Glycine	2.41±0.062 a	2.87±0.067 b	2.66±0.091 b	2.43±0.090 a	0.83±0.054	0.74±0.061	0.81±0.003	0.76 ± 0.03		
Serine	1.55±0.116 ab	1.39±0.041 a	1.73±0.002 b	1.68±0.111 b	0.75±0.003 a	0.66±0.033 b	0.74±0.013 a	0.67±0.004 b		
Alanine	3.35±0.109 a	3.38±0.094 ab	3.74±0.166 b	3.40±0.148 ab	0.80±0.05 a	0.72±0.046 a	0.99±0.029 b	0.79± 0.045 a		
Proline	2.94±0.106 ab	2.67± 0.020 a	2.98±0.143 ab	3.09± 0.174 b	1.12±0.043 a	1.06±0.063 a	1.88±0.029 b	1.13± 0.025 a		
Tyrosine	3.25±0.037 a	3.47±0.144 a	3.88±0.160 b	3.54±0.070 a	0.50±0.013 a	0.64±0.034 b	0.85±0.026 c	0.40±0.020 d		
Cystine	0.38±0.005 a	0.20±0.005 b	0.37±0.013 a	0.38±0.007 a	0.34±0.014 a	0.50±0.001 b	0.50±0.001 b	0.48±0.013 b		

a,b,c,d - means marked with different letters in the row (in the groups Larvae and Substrate separately) differed significantly (p<0.05, Fisher's LSD criterion). 1L—larvae (wheat bran + brewer's yeast + green potatoes); 2L—larvae (wheat bran + brewer's yeast + carrot). 3L—larvae (brewers' spent grain + brewer's yeast + carrot). 4CL—larvae, control (wheat bran + brewer's yeast + agar-agar gels); 1S—substrate (wheat bran + brewer's yeast + sprouted potatoes); 2S substrate (wheat bran + brewer's yeast + carrot). 3S—substrate (brewers' spent grain + brewer's yeast + carrot); 4CS—substrate, control (wheat bran + brewer's yeast + agar-agar gels):

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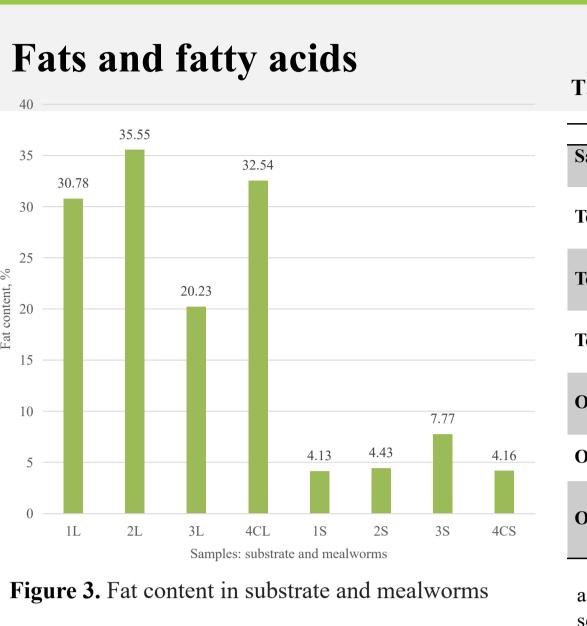


Table 3. Comparison of fatty acid in lyophilized larvae and substrate, % of total FAs content, average \pm standard error, n=3.

Fatty acids Samples	C12:0	C16:1	C17:0	C18:0	C18:1 cis	C18:2 trans	c18:3 gama	C20:1	C20:3 w6	C22:1 w9	C20:3w3	C22:4 w6	C20:5 w3
Larvae	2.46 ±1.411***	1.90 ±1.373***	3.39 ±3.821*	4.48 ±3.960*	35.72 ±2.472***	0.46 ±0.613***	0.62 ±0.504**	1.41 ±0.431***	1.58 ±1.341**	1.32 ±1.294**	4.14 ±5.674*	2.03 ±2.027 **	1.49± 1.371**
Substrate	0.11 ±0.201	0.07 ±0.132	0 ±0	1.75 ±0.51	18.12 ± 1.273	0 ±0	0.14 ± 0.233	7.16 ±2.084	0 ±0	0 ±0	0.53 ± 0.411	0.03 ±0.062	0.08 ± 0.135

In the groups larvae and substrate separately differed significantly (* - p<0,05; ** - p<0,01; *** - p<0,001, Fisher's LSD criterion); 1L-larvae (wheat bran + brewer's yeast + green potatoes); 2L-larvae (wheat bran + brewer's yeast + carrot). 3L-larvae (brewers' spent grain + brewer's yeast + carrot). 4CL-larvae, control (wheat bran + brewer's yeast + agar-agar gels); 1S-substrate (wheat bran + brewer's yeast + sprouted potatoes); 2S-substrate (wheat bran + brewer's yeast + carrot). 3S—substrate (brewers' spent grain + brewer's yeast + carrot); 4CS—substrate, control (wheat bran + brewer's yeast + agar-agar gels);

Main conclusions

- **%**).
- acids.
- demonstrated the highest content in 11 instances.
- *molitor* larvae for targeted applications.





Results

Table 4. Fatty acid ratio in lyophilized larvae and substrate, % of total FAs content, n=3.17

			Substrat	Δ				
			Lar	Substrate				
Samples	1L	2L	3L	4CL	1s	2s	3s	4cs
Fotal SFA								
	26.42 a	24.15 b	52.64 c	17.33 d	20.94 a	20.79 b	28.59 c	20.25 d
Fotal MUFA								
	45.12 a	38.02 b	35.89 c	43.37 d	24.64 a	28.47 b	24.13 c	24.18 d
Fotal PUFA								
	28.47 a	36.24 b	10.18 c	35.11 d	54.42a	50.74 b	44.63 c	54.05 d
Omega 6 FA								
	27.12 a	18.65 b	7.82 c	29.22 d	54.00 a	49.54 b	42.92 c	53.91 d
Omega 3 FA	0.22	4 20 1	1 (0 -	(00 1	0.02 -	0.201	2 (5 -	1 52 1
	0.32 a	4.39 b	1.69 c	6.99 d	0.03 a	0.29 b	2.65 c	1.53 d
Omega 6/3 FA								
	85.05 a	4.25 b	4.632 c	4.18 d	1838.83 a	170.51 b	16.20 c	35.34 d

a,b,c,d - means marked with different letters in the row (in the groups larvae and substrate separately) differed significantly (p<0.05, Fisher's LSD criterion); SFA – saturated fatty acids; MUFA - monounsaturated fatty acids; PUFA - polyunsaturated fatty acids;

The results showed that the highest protein content (59.18 ± 0.07 %) was observed in samples grown on brewers' spent grain, while wheat bran yielded the highest fat content (34.22 ± 0.491

The amount of FAs in the larvae was influenced by the substrate used, with variations in monounsaturated, omega-3, and oleic

Notably, wheat bran showed the highest content of total polyunsaturated FAs (36.23 %). In the analysis of 16 distinct amino acids, a sample with brewers' spent grain, consistently This adaptation renders the larvae suitable for diverse purposes, including animal or human nutrition and health enhancement. In conclusion, our study underscores the significance of

substrate optimization in harnessing the full potential of *Tenebio*